



Univerza v Mariboru

*Fakulteta za kemijo in  
kemijsko tehnologijo*



# Effect of high hydrostatic pressure on selected Black Yeast species

"Emerging Potential of Black Yeasts", 14-16 May 2010, Ljubljana



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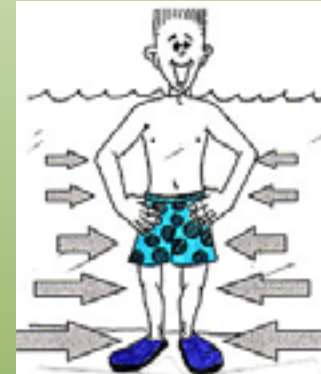
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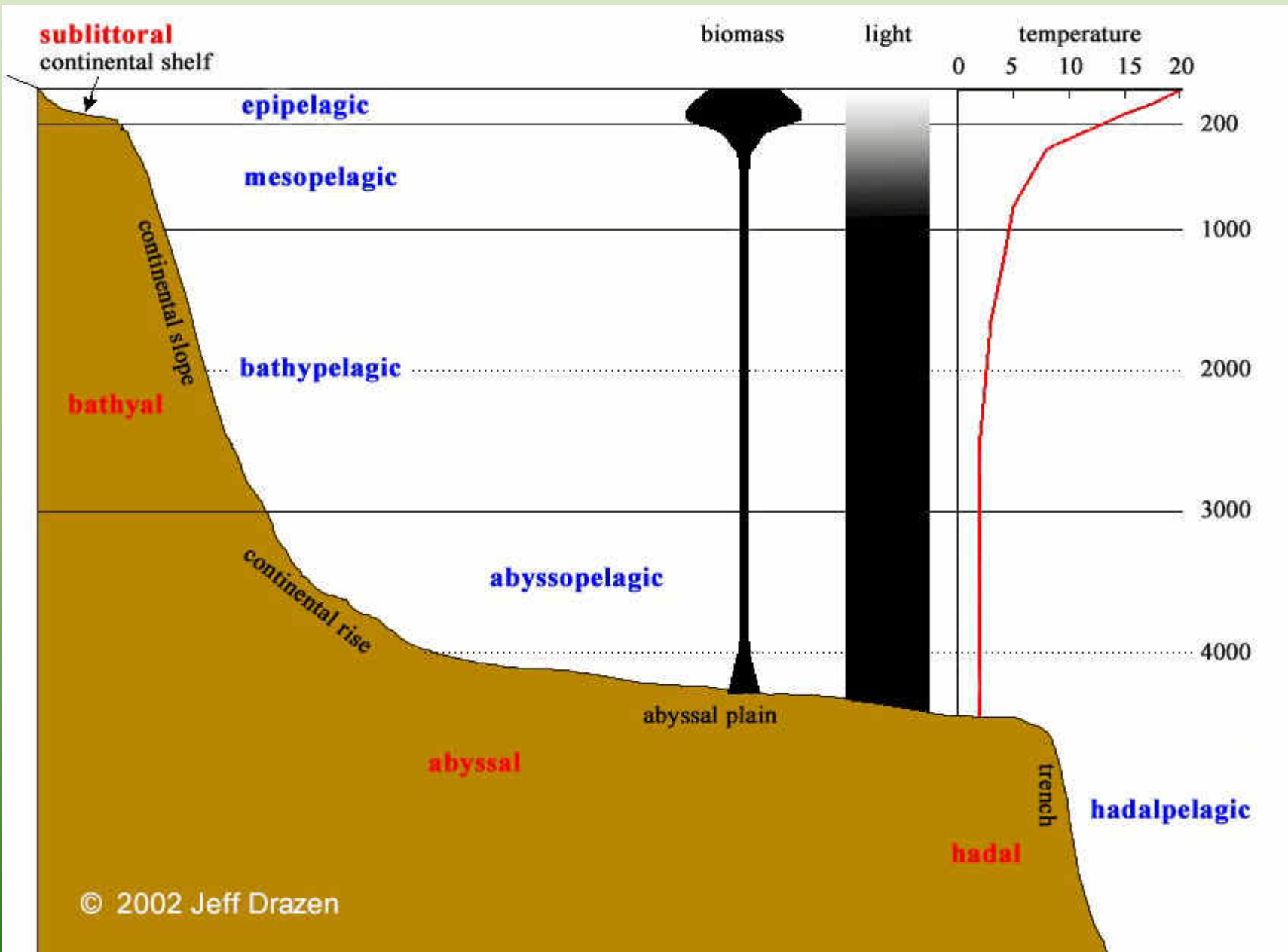
# Overview

- Introduction
  - hydrostatic pressure
  - microbial diversity
- Experimental design
- Results
- Conclusions and future directions

# Hydrostatic pressure

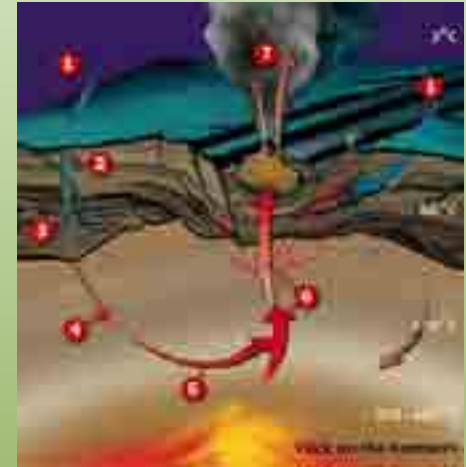
- Pressure increases with depth (oceans or underground)
- $1000\text{m} \text{---} 100\text{atm} = 101,325\text{bar} = 10\text{MPa}$
- Average depth of oceans  $3800\text{m}$  ( $38\text{MPa}$ ), maximum depth  $11000\text{m}$  ( $110\text{MPa}$ )
- Deep sea - water depth  $1000\text{m}$  and below
- High pressure biotopes: above  $100\text{MPa}$  pressure ( $10.000\text{m}$ )





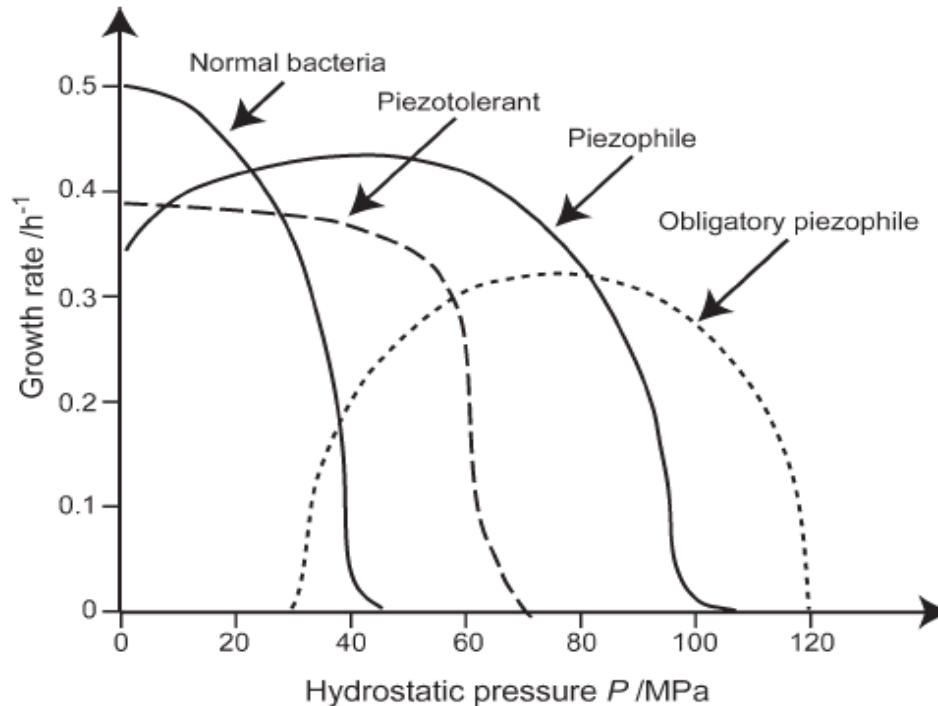
# High pressure biotopes and microbial diversity

- Until 19th century: no life below 600m
- 1940 high pressure biology (ZoBell and Morita)
- 1977 hidrothermal vents (2600m, invertebrae)
- 1979 first isolate of barophilic bacteria (Yayanos et al)
- 1993 first obligate barophilic anaerobic archaea (Erusto at al)
- 2009 **fungal diversity** in deep sea Hydrothermal Ecosystem (Le Calvez et al.)



<http://www.divediscover.whoi.edu/vents/vent-chemistry.html>

# Prefix "baro" = "piezo"



**Fig. 3** Definitions of the relations between growth rate of microorganisms and pressure. Piezophile microorganisms display a maximum growth at high pressure. They can either grow at atmospheric pressure or not, and are called strictly piezophile in the latter case. Piezotolerant microorganism grow best at atmospheric pressure, but can sustain high pressure, whereas mesophile or piezosensitive microorganisms totally stop growing at 40–50 MPa. (R. Margesin and Y. Nogi, *Cell. Mol. Biol.*, 2004, 50, 429)

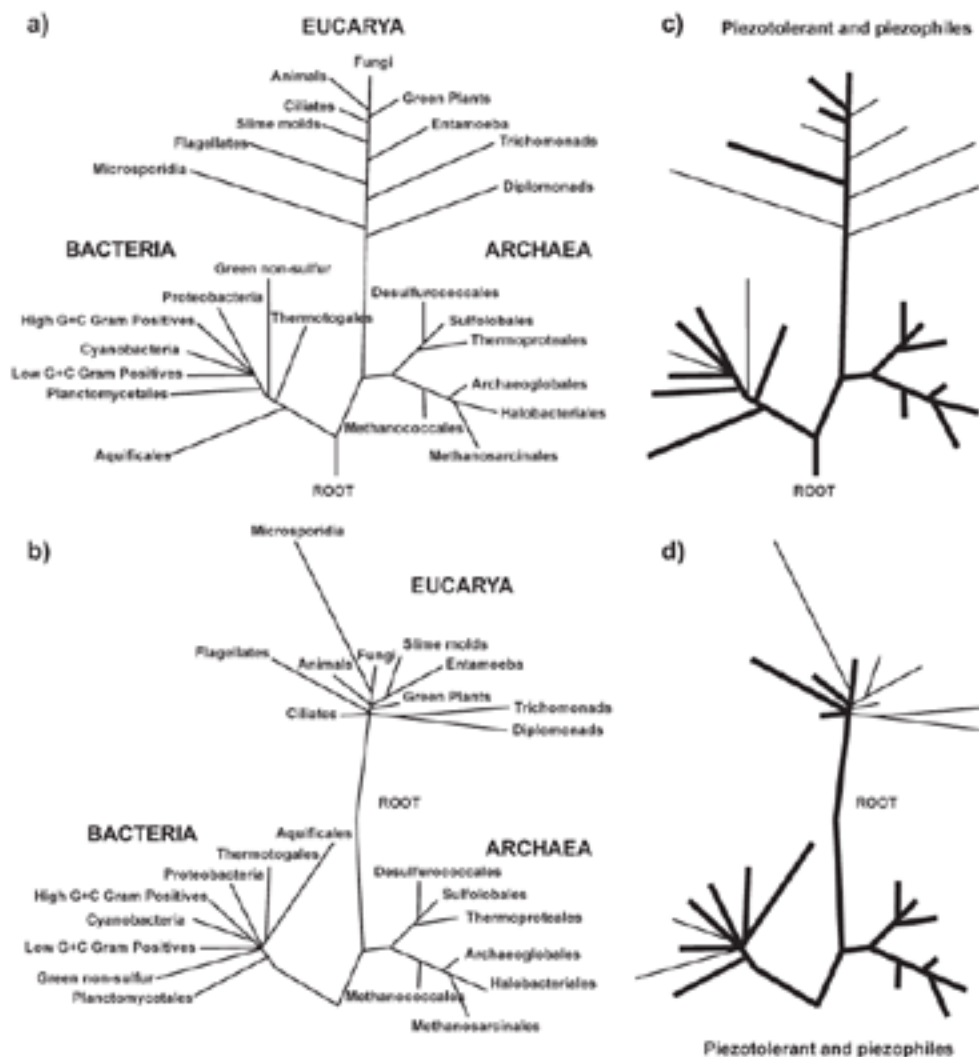


Fig. 2 Piezophily in the tree of life. (a) The classical view of the tree of life. The topology of the tree is mainly based on rDNA comparison. (b) A revised topology of the universal tree of life, after correction of phylogenetic pitfalls, such as long branch attraction or the removal of uninformative site from the analysis. (c) and (d) Distribution of piezophilic and piezotolerant organisms in the two trees of life. Thick black lines highlight bacterial groups in which piezotolerant and piezophile organisms have been characterized. Thin black lines highlight groups for which only piezotolerant species are known.

# Adapation mechanisms I

| BACTERIA & ARCHAEA                          | Source   | Depth (m)               | Adaptation  | Reference  |
|---|--|-------------------------|---|--|
| <i>Shewanella benthica</i>                  | Suruga Bay<br>Ryukyu Trench<br>Japan Trench<br>Izu-Bonin Trench<br>Ryukyu Trench | From<br>2485 to<br>6499 | All strains tested became more barophilic at higher temperatures.   | Kato C, Sato T, Horikoshi K: <b>Isolation and properties of barophilic and barotolerant bacteria from deep-sea mud samples.</b> <i>Biodiv Conserv</i> <b>1995</b><br>Kato C, Masui N, Horikoshi K: <b>Properties of obligatory barophilic bacteria isolated from a sample of deep-sea sediment from the Izu-Bonin trench.</b> <i>J Mar Biotechnol</i> <b>1996</b>                            |
| <i>Moritella japonica</i>                   | Japan Trench   | 6356                    |   | Li L, Kato C, Horikoshi K: <b>Distribution of the pressure regulated - operons in deep-sea bacteria.</b> <i>FEMS Microbiol Lett</i> <b>1998</b>  |
| DSK25 Gram-positive spore-forming bacterium | Japan Trench   | 6500                    |   |  |
| <i>Methanococcus jannaschii</i> (archaea)   |  |                         | Resistant proteases   | Michels PC, Clark DS: <b>Pressure-enhanced activity and stability of a hyperthermophilic protease from a deep-sea methanogen.</b> <i>Appl Environ Microbiol</i> <b>1997</b>  |
| Strain 2D2<br>Strain 16C1                   | intestinal contents of the deep-sea fish   | 6100<br>3100            | General shift from saturated to unsaturated fatty acids. Furthermore, these adaptive changes in response were also observed in response to low temperature. | Yutaka Yano, Akihiko Nakayama, Kenji Ishihara, and Hiroaki Saito <b>Adaptive Changes in Membrane Lipids of Barophilic Bacteria in Response to Changes in Growth Pressure.</b> <i>Appl Environ Microbiol</i> , February <b>1998</b>   |
| <i>Photobacterium profundum</i>             | Ryukyu Trench  |                         | Differences in respiratory chain (expression of certian cytochromes, amount of quinol oxidases)<br><br>Acumulation of protein-stabilizing solutes           | Qureshi, M.H., Kato, C., Horikoshi, K.,. <b>Purification of a cbb type quinol oxidase specifically induced in a deep-sea barophilic bacterium, <i>Shewanella sp. strain DB-172F.</i></b> <i>Extremophiles</i> <b>2 1998</b><br>Martin, D.D, Barlett, D.H, Roberts, M.F. <b>Solute accumulation in dep-sea bacterium <i>Photobacterium profundum.</i></b> <i>Extremophiles</i> , <b>2002.</b> |



Subject of detail gene expression profiling experiments



# Adaptation mechanisms II

| FUNGI   | Optimal growth properties (MPa) | Depth (m) | References   |
|---|---------------------------------|-----------|--|
| <i>Aspergillus ustus</i>  | 10                              | 860       | Raghukumar C, Raghukumar S. <b>Barotolerance of fungi isolated from deep-sea sediments of the Indian Ocean.</b> AQUATIC MICROBIAL ECOLOGY <b>1998.</b>   |
| <i>Graphium sp</i>  | 10                              | 965       |  |
| Debaryomyces hansenii, Rhodotorula rubra and Rhodosporidium sphaerocarpum | 20-40                           |           | LORENZ R. ; MOLITORIS H. P. ; <b>Cultivation of fungi under simulated deep sea conditions.</b> Mycological research , <b>1997</b>  |
| 181 cultures of fungi (25 representative )                                | 20 (30)                         | 5000      | Damare S, Raghukumar C, Raghukumar S. <b>Fungi in deep-sea sediments of the Central Indian Basin.</b> Deep-sea research part i-oceanographic research papers, <b>2006</b>  |
| deep-sea <i>Aspergillus</i> isolates                                      | 20                              | 5000      | Samir R. Damare-Manju Nagarajan and Chandralata Raghukumar <b>Spore germination of fungi belonging to <i>Aspergillus</i> species under deep-sea conditions .</b> In Deep Sea Research Part I: Oceanographic Research Papers, <b>2008</b> |
| <i>Saccharomyces cerevisiae</i> (MODEL)                                   | 18                              |           | Damare S, Raghukumar C, Raghukumar S <b>Effect of temperature on the role of Hsp104 and trehalose in barotolerance of <i>Saccharomyces cerevisiae</i>.</b> FEBS Letters, <b>1997</b>   |



- morphology  
- ability to germinate

# Effect of high pressure on *S. cerevisiae*

F. Abe, *Cell. Mol. Biol.*, 2004, 50, 437.

Table 1 Pressure effects on yeast physiology<sup>30</sup>

| Pressure/MPa | Effects   |
|--------------|---|
| 0.1–50 MPa   | Arrest of cell growth<br>Metabolic changes<br>Inhibition of amino acid uptake<br>Stress-inducible expression              |
| 50 MPa–      | Inhibition of ethanol fermentation<br>Internal acidification<br>Stress-inducible expression                               |
| 100 MPa–     | Reduction in viability<br>Membrane, and cell wall perturbations<br>Acquired piezotolerance<br>Stress-inducible expression |
| 200 MPa–     | Alteration of genome preservation<br>Shrinkage and leakage of cells<br>Stress-inducible expression                        |

- **On gene expression:**
  - Up-regulation of genes involved in stress tolerance and carbohydrate metabolism
  - Down-regulation of genes involved in cellular transcription, protein synthesis and cell cycle
- **On protein synthesis:**
  - RNA synthesis is maintained
  - Improved piezoresistance AFTER being exposed to a mild stress
- **On fluidity of the membrane:**
  - Increase of unsaturated fatty acids
- **On cell wall and cytoskeleton:**
  - Up-regulation of Hsp12p in cell wall, where it was proposed that acts as a plasticizer
- **Intracellular changes:**
  - Acidification of cytoplasm and vacuole

Time of exposure to HP is important factor!  
(50MPa for 24hrs results in 100% mortality)

HP treatment of food products (300-500MPa)

# Multiple response of black yeasts to stress

- production of melanin  
-osmoadaptation



- Meristematic/microcolonial growth (keeping the optimal volume-to-surface ratio)

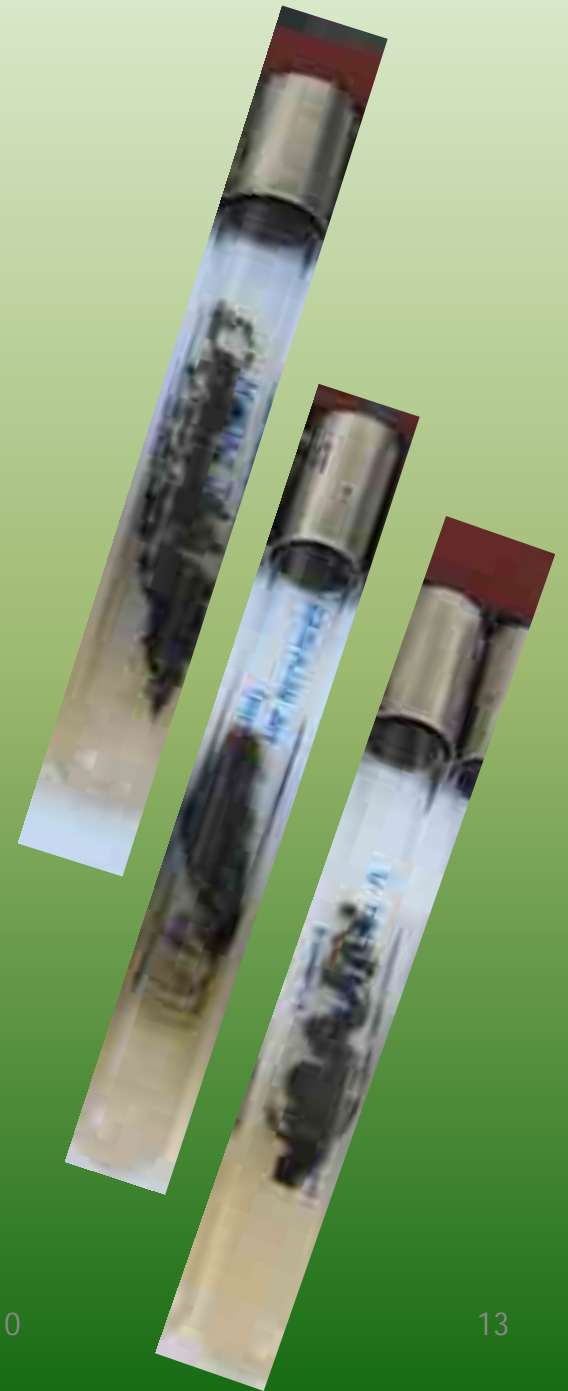


# Studied fungi

|     |   |            |                |
|-----|---|------------|----------------|
| 1.  | <i>Hortaea werneckii</i>                        | EXF - 225  | Salterns       |
| 2.  | <i>Trimmatostroma salinum</i>                   | EXF - 295  | Salterns       |
| 3.  | <i>Phaeotheca triangularis</i>                  | EXF-206    | Salterns       |
| 4.  | <i>Cladosporium halotolerans</i>                | EXF - 572  | Salterns       |
| 5.  | <i>Cladosporium spinulosum</i>                  | EXF - 334  | Salterns       |
| 6.  | <i>Candida parapsilosis</i>                     | EXF - 517  | Salterns       |
| 7.  | <i>Pichia guillermondii</i>                     | EXF - 1496 | Arctic         |
| 8.  | <i>Pichia guillermondii</i>                     | EXF - 518  | Salterns       |
| 9.  | <i>Aureobasidium pullulans var. pullulans</i>   | EXF-150    | Salterns       |
| 10. | <i>Aureobasidium pullulans var. pullulans</i>   | EXF-1668   | Arctic         |
| 11. | <i>Aureobasidium pullulans var. melanogenum</i> | EXF - 924  | Arctic         |
| 12. | <i>Aureobasidium pullulans var. melanogenum</i> | EXF - 3382 | Deep sea       |
| 13. | <i>Aureobasidium pullulans var. subglaciale</i> | EXF - 2481 | Arctic         |
| 14. | <i>Aureobasidium pullulans var. subglaciale</i> | EXF-2479   | Arctic         |
| 15. | <i>Aureobasidium pullulans var. namibiae</i>    | EXF - 3398 | Namibia        |
| 16. | <i>Hormonema 1</i>                              | EXF - 914  | Arctic         |
| 17. | <i>Hormonema 2</i>                              | EXF - 2500 | Arctic         |
| 18. | <i>Cryptococcus liquefaciens</i>                | EXF-3584   | Arctic         |
| 19. | <i>Rhodotorula mucilaginosa</i>                 | EXF 1630   | Arctic         |
| 20. | <i>Rhodotorula mucilaginosa</i>                 | EXF-3790   | Salterns       |
| 21. | <i>Cryptococcus diffluens</i>                   | EXF-3772   | Salterns       |
| 22. | <i>Cryptococcus albidus</i>                     | EXF-3477   | Arctic         |
| 23. | <i>Penicillium crustosum</i>                    | EXF 763    | Salterns       |
| 24. | <i>Penicillium crustosum</i>                    | EXF-1329   | Arctic, CREA + |
| 25. | <i>Penicillium crustosum</i>                    | EXF-1005   | Arctic, CREA - |
| 26. | <i>Debaryomyces hansenii</i>                    | EXF-589    | Salterns       |
| 27. | <i>Debaryomyces hansenii</i>                    | EXF-1590   | Arctic         |
| 28. | <i>Wallemia ichthyophaga</i>                    | EXF-994    | Salterns       |
| 29. | <i>Wallemia muriae</i>                          | EXF-951    | Salterns       |
| 30. | <i>Wallemia sebi</i>                            | EXF-958    | Pumpkin seed   |
| 31. | <i>Saccharomyces cerevisiae</i>                 | EXF-531    | control        |

# Selected black yeasts under hydrostatic pressure

- *Trimmatostroma salinum*  
EXF – 295, Solar salterns
- *Aureobasidium pullulans* var. *melanogenum*  
EXF – 924, Arctic ice
- *Hormonema 1*  
EXF – 914, Arctic ice
- *Saccharomyces cerevisiae* (positive control)



# Experimental design

7 days of growth



MEA

10 days of growth



MEA + 10% NaCl



Cells in 0,9%NaCl solution



Cells in 10 %NaCl solution



*immediate  
pressurization*



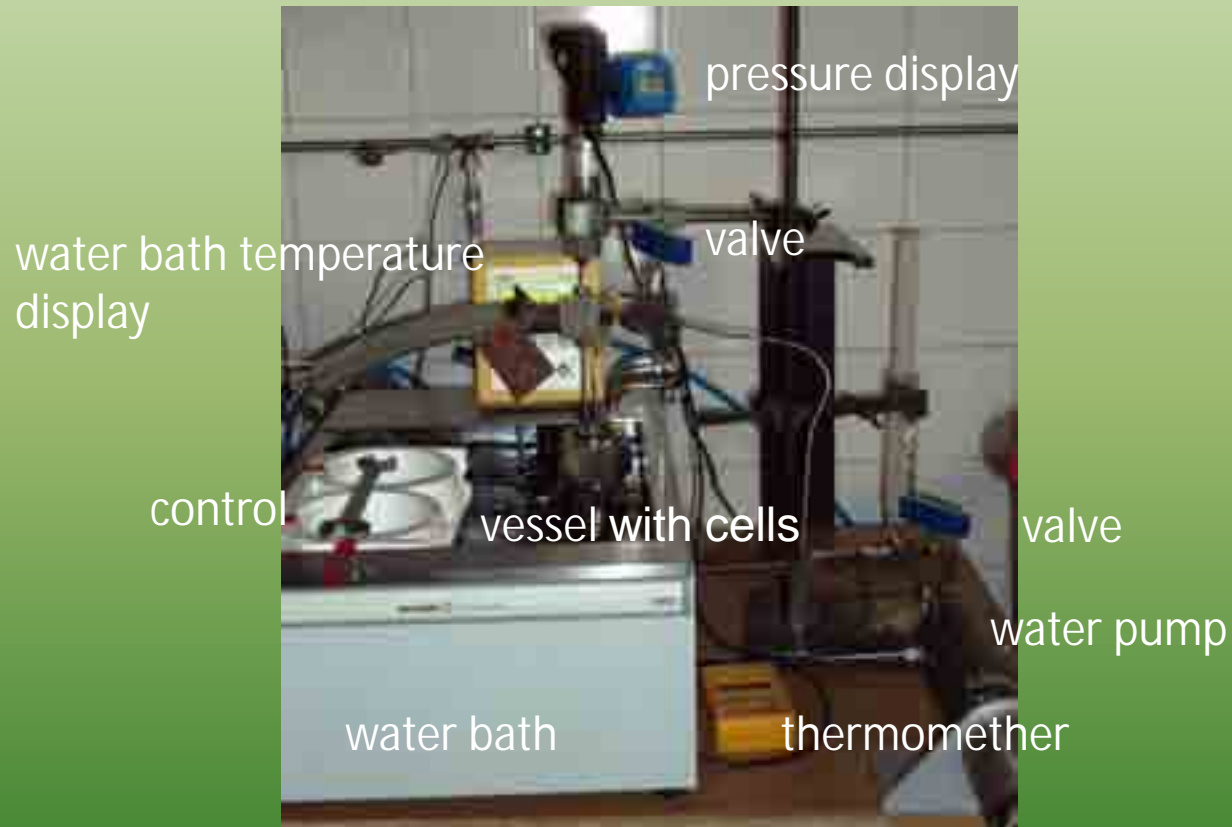
High pressure vessel  
24hrs, 300 bar (30MPa) room  
temperature (23°C)

MEA



MEA + 10% NaCl

# High pressure equipment



High pressure equipment (UniMb)

# Results

## *Trimmatostroma salinum*

MEA

MEA + 10% NaCl

control

after exposure to HP

control

after exposure to HP



↓ number



↓ number

↑ colony diameter



# Results

## *Aureobasidium pullulans* var. *melanogenum*

### MEA

control

after exposure to HP



### MEA + 10% NaCl

control

after exposure to HP



No differences observed

↓ number  
↑ colony diameter

# Results

## *Hormonema 1*

MEA

MEA + 10% NaCl

control

after exposure to HP

after exposure to HP

control



No differences observed



number



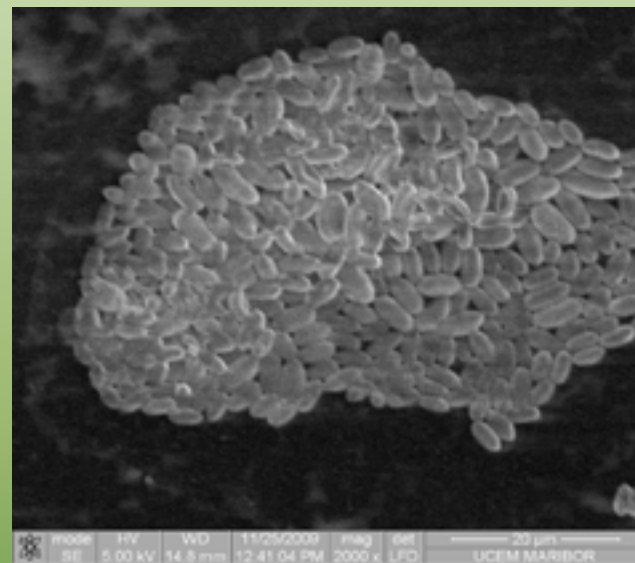
colony diameter

# SEM

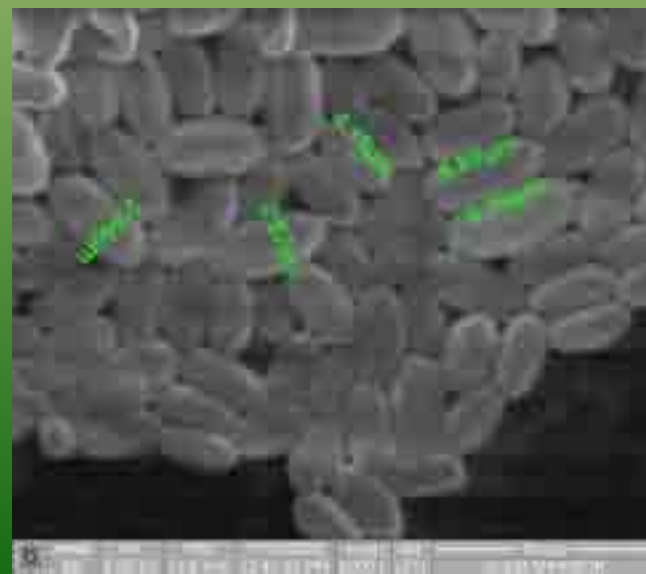
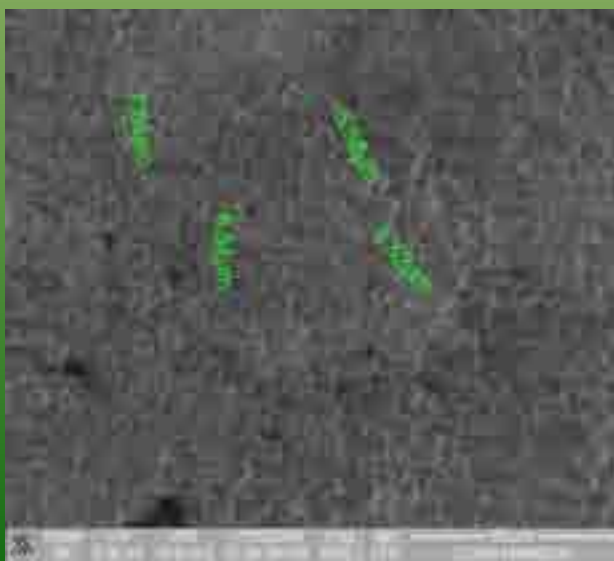
## *Hormonema 1*

control

after exposure to HP



MEA



Emerging potential of Black  
Yeasts, 14-16 May 2010

# SEM

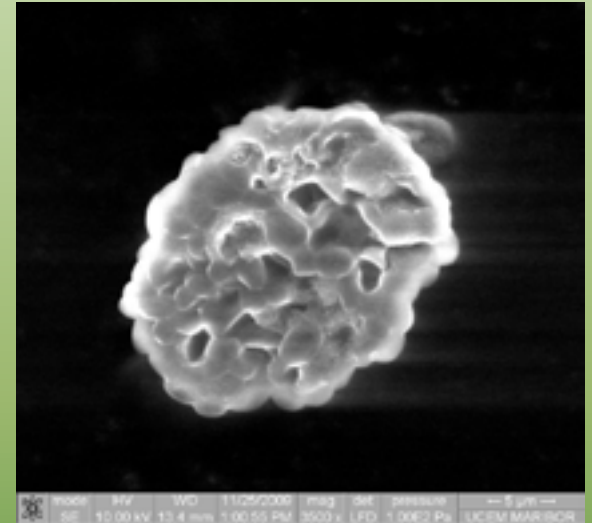
## *Hormonema 1*

control



MEA + 10%NaCl

after exposure to HP



Clumps:

- High concentration of salt
- HP

# Results

## *Saccharomyces cerevisiae*

### MEA

after exposure to HP

control



### MEA + 10% NaCl

after exposure to HP

control



# Conclusions and future directions

- Cells of selected black yeasts are able to survive exposure to 30 MPa
- Exposure to 30MPa pressure can cause changes in colony appearance
- Ability of subglacial *Hormonema* to adjust volume to surface ratio (long and narrow shape of cells after exposure to HP; SEM)
- Future plan - increase of hydrostatic pressure!!!

# Thank you for your attention!

